# EUCAST v12.0 Breakpoints for Piperacillin/Tazobactam on MicroScan Dried Gram Negative MIC Panels for Enterobacterales species and Pseudomonas species

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### **ABSTRACT**

Background: MIC data from MicroScan Dried Gram-negative MIC (MSDGN) Panels with piperacillin/tazobactam were evaluated with EUCAST v12.0 breakpoints from a multicenter clinical study. MIC results from Enterobacterales and Pseudomonas spp were compared to results obtained with CLSI frozen broth microdilution panels.

Material/methods: The study included a total of 547 Enterobacterales and 85 Pseudomonas spp. clinical isolates tested using the turbidity and Prompt methods of inoculation during the combined phases of efficacy and challenge. MSDGN panels were evaluated at three clinical sites by comparing MIC values obtained using the MSDGN panels to MICs utilizing a CLSI broth microdilution reference panel. MSDGN panels were incubated at 35 ± 1°C and read on the WalkAway System, the autoSCAN-4 instrument, and read visually at 16-20 hours. Frozen reference panels were prepared according to CLSI/ISO methodology, incubated for 16-20 hours for Enterobacterales and Pseudomonas spp. and read visually. EUCAST v12.0 breakpoints (mg/L) used for interpretation of MIC results were:  $\leq 8/4$  S, > 8/4 R for Enterobacterales and  $\leq 0.001/4$  S, > 16/4 R for Pseudomonas spp.

Results: When compared to frozen reference panel results, essential and categorical agreements for all isolates tested in efficacy and challenge are as follows (AS-4 read method yielded similar results) with the following recommendation: Due to low performance for S. marcescens and piperacillin/tazobactam with WalkAway read method with Prompt inoculation, results should be confirmed by a manual read prior to reporting. Due to the performance with P. rettgeri, S. liquefaciens, and S. liquefaciens complex and all read and inoculation methods, do not report drug, therapy, or MIC:

Organism Group		Essential nent* %	1 ' " 1		Prompt Errors		Prompt Very Major Error** %	
Огоир	WalkAway	Manual	WalkAway	Manual	WalkAway	Manual	WalkAway	Manual
Enterobacterales	93.4 (510/546)	96.5 (527/546)	94.5 (516/546)	97.6 (533/546)	3.8 (17/452)	0.4 (2/452)	1.1 (1/94)	1.1 (1/94)
Pseudomonas species	94.1 (80/85)	95.3 (81/85)	96.5 (82/85)	96.5 (82/85)	3.2 (2/63)	1.6 (1/63)	4.6 (1/22)	4.6 (1/22)
All Organisms***	93.7 (638/681)	96.3 (656/681)	94.8 (598/631)	97.5 (615/631)	3.7 (19/515)	0.6 (3/515)	1.7 (2/116)	1.7 (2/116)
*Overall EA palavieted for all assentance averall CA palavieted for all assentance with interpretive evitoric								

\*Overall EA calculated for all organisms, overall CA calculated for all organisms with interpretive criteria \*Calculation excluding 1 well errors, Pseudomonas spp. errors are potential errors \*\*\* All organisms include Acinetobacter spp., Enterobacterales, and Pseudomonas spp. organism groups

Conclusions: Piperacillin/tazobactam MIC results for Enterobacterales and Pseudomonas spp. obtained with the MSDGN panel correlate well with MICs obtained using frozen reference panels with updated EUCAST v12.0 interpretive criteria in this multicenter study.

### INTRODUCTION

A multicenter study was performed to evaluate the performance of a MicroScan Dried Gram Negative MIC panel with piperacillin/tazobactam using Enterobacterales and Pseudomonas spp. isolates with EUCAST interpretive breakpoints.

### **METHODS**

#### Study Design

MSDGN MIC panels were tested concurrently with a CLSI frozen broth microdilution reference panel at three sites using both the turbidity and Prompt inoculation methods on a total of 683 Acinetobacter spp., Enterobacterales, and *Pseudomonas* spp. clinical isolates.

Quality Control Expected Results (CLSI M100 ED33)

Escherichia coli ATCC 25922: 1/4 - 8/4 µg/ml Pseudomonas aeruginosa ATCC 27853: 1/4 - 8/4 µg/ml Escherichia coli ATCC 35218: 0.5/4 - 2/4 µg/ml

### **METHODS** (Continued)

#### **Panels**

•Frozen reference and MSDGN MIC panels contained two-fold doubling dilutions of piperacillin/tazobactam 8/4-256/4 µg/ml in cation-adjusted Mueller-Hinton broth. Reference panels were prepared and frozen following CLSI/ISO recommendations.

#### Reproducibility

•Reproducibility organisms with known results on-scale for piperacillin/tazobactam were tested in triplicate (for each inoculation method) on the MSDGN MIC panels and singly on the frozen reference panel on three different days at each site.

•MSDGN MIC panels were tested using both the turbidity and Prompt inoculation methods and read on the WalkAway system, autoSCAN-4 instrument, and visually.

#### **Quality Control**

•Quality control (QC) testing was performed daily using ATCC 25922 Escherichia coli, ATCC 27853 Pseudomonas aeruginosa, ATCC 35218 Escherichia coli with CLSI QC ranges.

#### Panel Inoculation, Incubation, and Reading

•All isolates were subcultured into trypticase soy agar (TSA) with 5% sheep blood and incubated for 18-24 hours at 35-37°C prior to testing. Isolates from frozen stocks were subcultured twice before testing.

•Inoculum suspensions for each strain were prepared with the direct standardization (turbidity standard) method for MSDGN MIC and frozen reference panels. MSDGN MIC panels were also inoculated using the Prompt Inoculation method.

•Following inoculation, MSDGN MIC panels were also incubated at  $35\pm1^{\circ}$ C in the WalkAway system for  $18\pm2$  hours. All panels were read by the WalkAway, autoSCAN-4 and visually.

#### Data Analysis

•Essential Agreement (EA) = MSDGN panel MIC within +/- 1 dilution of the frozen reference result MIC.

•Categorical Agreement (CA) = MSDGN panel and reference categorical results (S and R) agree using EUCAST v12.0 breakpoints for Enterobacterales and Pseudomonas species. Acinetobacter species were evaluated for MIC reporting only (Table 1). Due to arbitrary, "offscale" breakpoint of ≤ 0.001 for *Pseudomonas* species, in this analysis, S = I (Susceptible, increased exposure" (I)), so all errors are calculated as potential major errors and potential very major errors.

Table 1. Piperacillin/Tazobactam EUCAST Breakpoints (ug/ml) EUCAST v12.0 is equivalent to EUCAST v13.0 interpretive criteria

Organism Group	S	R				
Acinetobacter spp.	IE	IE				
Enterobacterales	≤ 8/4	> 8/4				
Pseudomonas spp.	≤ 0.001/4	> 16/4				
PK/PD	≤ 8/4	> 16/4				
Vibrio Spp.	≤ 1/4	> 1/4				

•Major Errors = Frozen reference MIC is S and MSDGN panel MIC is R; calculated for susceptible strains only. No. Major Errors

% Major Errors =

Total No. S Isolates tested •Very Major Errors = Frozen reference is R and MSDGN panel MIC is S; calculated for resistant strains only.

No. Very Major Errors

% Very Major Errors = - X 100 Total No. R Isolates tested

•Minor Errors = Frozen reference is S or R when MSDGN panel MIC is I or MSDGP panel MIC is S or R when frozen reference is I; calculated for all isolates tested. No. Minor Errors

> % Minor Errors = Total No. Isolates tested

- X 100

## **RESULTS**

Efficacy (Tables 2 and 3)

•A total of 683 Acinetobacter spp., Enterobacterales and Pseudomonas spp. clinical isolates were tested among three sites. MSDGN panels were inoculated using the turbidity inoculation method. Essential Agreement for Acinetobacter spp., Enterobacterales and Pseudomonas spp. between MSDGN panel and frozen reference panel was 98.0% (669/683) for manual read method, 96.1% (656/683) for WalkAway System, 96.6% (660/683) for autoSCAN-4 instrument using the turbidity inoculation method.

•Categorical Agreement for Enteropacterales and Pseudomonas spp. between MSDGN panel and frozen reference panel was 96.8% (613/633) for manual read method, 95.9% (607/633) for WalkAway System, 96.7% (612/633) for autoSCAN-4 instrument using the turbidity inoculation method.

Table 2. Efficacy - Turbidity Inoculation Method

	Essential		Categorical		Major		Very Major	
	Agreement		Agreement Er		Err	ors	Errors	
<b>Read Method</b>	No.	%	No.	%	No.	%	No.	%
Manual	669/683	98.0	613/633	96.8	0/517	0.0	2/116	1.7
WalkAway	656/683	96.1	607/633	95.9	4/517	0.8	2/116	1.7
autoSCAN-4	660/683	96.6	612/633	96.7	0/517	0.0	2/116	1.7

•A total of 681 Acinetobacter spp., Enterobacterales and Pseudomonas spp. clinical isolates were tested among three sites. MSDGN panels were inoculated using the Prompt inoculation method. Differences in Prompt and turbidity totals tested are due to smaller, pin-point colonies not suitable for Prompt inoculation per instructions in the Prompt procedural manual.

•Essential Agreement for Acinetobacter spp., Enterobacterales and Pseudomonas spp. between MSDGN panel and frozen reference panel was 96.3% (656/681) for manual read method, 93.7% (638/681) for WalkAway System, 95.9% (653/681) for autoSCAN-4 instrument using the Prompt inoculation method.

•Categorical Agreement for Enterobacterales and Pseudomonas spp. between MSDGN panel and frozen reference panel was 97.5% (615/631) for manual read method, 94.8% (598/631) for WalkAway System, 97.0% (612/631) for autoScan-4 instrument using the Prompt inoculation method.

Table 3. Efficacy - Prompt Inoculation Method

	Essential		Catego	orical Ma		jor	Very Major	
	Agreement		Agreement		Errors		Errors	
<b>Read Method</b>	No.	%	No.	%	No.	%	No.	%
Manual	656/681	96.3	615/631	97.5	3/515	0.6	2/116	1.7
WalkAway	638/681	93.7	598/631	94.8	19/515	3.7	2/116	1.7
autoSCAN-4	653/681	95.9	612/631	97.0	3/515	0.6	2/116	1.7

Performance problems with P. rettgeri, S. liquefaciens and S. liquefaciens complex resulted in decisions to not report the drug, therapy, or MIC. Results for S. marcescens with WalkAway read method with Prompt inoculation, should be confirmed by a manual read prior to reporting because of elevated major error rates for WalkAway reads with Prompt inoculation.

Reproducibility (Tables 4 & 5)

•Overall agreement (within  $\pm$  two-fold dilution) between all sites for the reproducibility phase was ≥ 95% for all combinations.

Table 4. Reproducibility Best Case - All Sites combined

Read Method	Inoculation Method	No. (%) Agreement Best Case All Sites Combined				
Manual		262/270 (97.0)				
WalkAway	Turbidity	y 258/270 (95.6)				
autoSCAN-4		258/270 (95.6)				
Manual		260/270 (96.3)				
WalkAway	Prompt	Best Case All Sites Combined 262/270 (97.0) 258/270 (95.6) 258/270 (95.6)				
autoSCAN-4		263/270 (97.4)				

Table 5. Reproducibility Worst Case - All Sites combined

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Read Method	Inoculation Method	No. (%) Agreement Worst Case All Sites Combined				
Manual		262/270 (97.0)				
WalkAway	Turbidity	258/270 (95.6)				
autoSCAN-4		258/270 (95.6)				
Manual		260/270 (96.3)				
WalkAway	Prompt	263/270 (97.4)				
autoSCAN-4		263/270 (97.4)				

Quality Control (Tables 6 and 7)

Overall quality control results were >95% for each read and inoculation method on the dried test panel for ATCC 25922 E. coli, ATCC 27853 P. aeruginosa and ATCC 35218 E. coli. Quality control results were >95% for the frozen reference panel, which were read manually with turbidity inoculation method. The number of replicates and percentage within range are indicated in Tables 6 and 7. Variations in total number tested for each read method are due to technical error elimination.

Table 6. Quality Control - Dried Test Results

		Percent (%) in Range					
0	QC Range	Manual		WalkAway		autoSCAN-4	
Organism	(µg/mL)	Turbidity	Prompt	Turbidity	Prompt	Turbidity	Prompt
E. coli	1/4 – 8/4	98/98	98/98	98/98	98/98	98/98	98/98
ATCC 25922	1/4 - 0/4	100%	100%	100%	100%	100%	100%
P. aeruginosa	1/4 – 8/4	95/98	95/97	95/98	95/97	96/98	95/97
ATCC 27853	1/4 - 6/4	96.9%	97.9%	96.9%	97.9%	98.0%	97.9%
E. coli	0.5/4 - 2/4	98/98	96/96	98/98	96/96	98/98	96/96
ATCC 35218	0.5/4 - 2/4	100%	100%	100%	100%	100%	100%

Table 7. Quality Control - Frozen Reference Results

Ormaniam	QC Range	Manual
Organism	(µg/mL)	Turbidity
E. coli	1/4 - 8/4	100%
ATCC 25922	(frozen)	(98/98)
P. aeruginosa	1/4 - 8/4	98.0%
ATCC 27853	(frozen)	(96/98)
E. coli	0.5/4 - 2/4	100%
ATCC 35218	(frozen)	(98/98)

### CONCLUSION

There is a strong correlation between the MIC results obtained using MicroScan Dried Gram Negative panel and MICs obtained using a CLSI broth microdilution frozen reference panel for susceptibility testing of piperacillin/tazobactam and Acientobacter spp.. Enterobacterales and Pseudomonas spp. in a multicenter study using EUCAST v12.0 interpretive criteria.

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